LETTERS

Rare chromosomal deletions and duplications increase risk of schizophrenia

The International Schizophrenia Consortium*

Schizophrenia is a severe mental disorder marked by hallucinations, delusions, cognitive deficits and apathy, with a heritability estimated at 73-90% (ref. 1). Inheritance patterns are complex, and the number and type of genetic variants involved are not understood. Copy number variants (CNVs) have been identified in individual patients with schizophrenia²⁻⁷ and also in neurodevelopmental disorders⁸⁻¹¹, but large-scale genome-wide surveys have not been performed. Here we report a genome-wide survey of rare CNVs in 3,391 patients with schizophrenia and 3,181 ancestrally matched controls, using high-density microarrays. For CNVs that were observed in less than 1% of the sample and were more than 100 kilobases in length, the total burden is increased 1.15-fold in patients with schizophrenia in comparison with controls. This effect was more pronounced for rarer, single-occurrence CNVs and for those that involved genes as opposed to those that did not. As expected, deletions were found within the region critical for velo-cardio-facial syndrome, which includes psychotic symptoms in 30% of patients¹². Associations with schizophrenia were also found for large deletions on chromosome 15q13.3 and 1q21.1. These associations have not previously been reported, and they remained significant after genome-wide correction. Our results provide strong support for a model of schizophrenia pathogenesis that includes the effects of multiple rare structural variants, both genome-wide and at specific loci.

The International Schizophrenia Consortium was established to promote rapid progress towards the identification of genetic causes underlying schizophrenia. The consortium is composed of investigators from the University of Aberdeen, Cardiff University, the University of Edinburgh, Karolinska Institutet, Massachusetts General Hospital, the University of North Carolina-Chapel Hill, the Queensland Institute of Medical Research, the University of Southern California, the Stanley Center for Psychiatric Research at the Broad Institute of Harvard and MIT, Trinity College Dublin and University College London.

We surveyed single nucleotide polymorphisms (SNPs) and CNVs using the Affymetrix Genome-Wide Human SNP 5.0 and 6.0 arrays in European cases of schizophrenia and in ancestrally matched controls (Table 1 and Supplementary Information)¹³. On the basis of the genome-wide SNP data there was no evidence of major population stratification within each site¹⁴ (data not shown). Intensity data from both SNP and CNV probes were used to identify autosomal deletions and duplications, based on a hidden Markov model¹⁵.

This study focused on rare but highly penetrant structural variation in schizophrenia, following a natural extension of the classical medical genetic approach. Common CNVs are better identified with different algorithms and are better tested for association separately^{13,15}. Considering CNVs that were present in less than 1% of our total sample, there were 6,753 larger than 100 kilobases (kb) that passed sample and CNV quality filtering (see Supplementary Information and Supplementary Table 1). The median size was 182.1 kb (166.3 kb for deletions, 194.4 kb for duplications), 39% were deletions and the median number per individual was 1. We assessed the impact of rare structural variation on the risk for schizophrenia in two ways: first in terms of an individual's genome-wide burden, and second by searching for specific loci that were significantly associated with disease.

Structural variants have been identified for severe neurodevelopmental disorders^{9–11,16,17}. Because it has been postulated that schizophrenia might, at least in part, have a developmental aetiology¹⁸, we posited a role for CNVs in schizophrenia, as have others^{2–6}. Several loci have been identified, including variants containing genes with neurodevelopmental roles^{2–5}. However, a critical question is the extent to which this is a general mechanism for producing schizophrenia in typical clinical populations rather than in cases selected for atypical phenotypic features such as very early onset or mental retardation. This motivated our primary hypothesis: that individuals with schizophrenia have a greater genome-wide burden of CNVs. Considering all CNVs, we observed that cases had a greater average burden than controls (one-sided, empirical $P = 3 \times 10^{-5}$ controlling for array type; Table 2). Controls on average had 0.99 CNVs per person, whereas cases showed a 1.15-fold higher rate.

We next explored this subtle, but highly statistically significant, observation of increased burden. We defined burden in two ways: as the number of CNVs carried by an individual (as above), and also as the number of genes spanned by those CNVs. This second metric (the 'gene count') in fact showed a stronger association with schizophrenia (1.41-fold increase, empirical $P = 2 \times 10^{-6}$) than burden defined simply as the number of CNVs. Characteristics of CNV subgroups studied here are their frequency, type, size, and proximity to a gene (Tables 2 and 3, and Supplementary Table 2). We observed an increased burden across multiple independent subgroups of CNVs,

Table 1	Study	sample	characteristics	and ger	otyping	platforms
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Sample	Ancestry	Case (n)	Control (n)	Genotyping platform				
University of Aberdeen	Scottish	727	694†	5.0				
University College London	British	547	n/a‡	5.0				
Portuguese Island Collection	Portuguese	333	200†	5.0				
Karolinska Institutet	Swedish	622	437	5.0/6.0§				
Cardiff University	Bulgarian	479*	646	6.0				
Trinity College Dublin	Irish	280	914	6.0				
University of Edinburgh	Scottish	403*	290	6.0				

Figures are the numbers of cases and controls passing quality control and included in the final analyses. Case samples received a diagnosis of schizophrenia. 'Genotyping platform' indicates Affymetrix array type (5.0 or 6.0).

* Cases were excluded if IQ was less than 70.

† Controls were screened for psychiatric disorders.

‡ University College London control samples genotyped with the Affymetrix 500K two-chip genotyping platform were excluded because CNV data were not available.

§ Swedish cases and controls matched for array type for all analyses.

Table 2 | Global CNV burden analysis: event type and frequency

	Frequency		CNV burden (number)			CNV burden (gene count)		
CNV type		CNV (n)	Р	Case/control ratio	Baseline rate (controls)	Р	Case/control ratio	Baseline rate (controls)
Deletions and duplications	All	6,753	3×10^{-5}	1.15	0.99	2×10^{-6}	1.41	2.01
	Single occurrence	890	5×10^{-6}	1.45	0.11	0.0057	1.67	0.32
	2-6 occurrences	2,465	0.0013	1.16	0.35	5×10^{-4}	1.36	0.80
Deletions only	All	2,652	0.11	1.08	0.40	3×10^{-5}	1.55	0.72
	Single occurrence	470	0.011	1.29	0.06	0.005	1.77	0.12
	2-6 occurrences	994	0.048	1.15	0.15	0.13	1.38	0.21
Duplications only	All	4,101	2×10^{-5}	1.20	0.59	10^{-4}	1.28	1.94
	Single occurrence	734	8×10^{-6}	1.58	0.09	0.015	1.60	0.30
	2-6 occurrences	1,532	0.011	1.16	0.22	0.012	1.30	0.69

The table shows an analysis of global CNV burden in cases versus controls. As described in the text, CNVs have previously been filtered for a maximum of about 1% sample frequency. These analyses were further stratified according to type (deletions versus duplications) and frequency (single occurrences and CNVs observed two to six times). Empirical *P* values (one-sided, controlling for array type) are given for two measures of CNV burden (number of CNVs and number of genes affected by CNVs). The average rate in controls (baseline, number of CNVs per person) and the fold increase in case (case/control ratio) are shown for each analysis. Note that the 'Deletions only' and 'Duplications only' counts are not expected to sum to the 'Deletions and duplications' count for the two lower-frequency groups (see Supplementary Information).

a finding that was more pronounced for rarer CNVs and those involving genes. Deletions and duplications also had somewhat different profiles: the association of deletions varied more noticeably with respect to CNV size and proximity to a gene, whereas duplications showed a more uniform pattern.

A total of 890 CNVs were observed in either a case or a control as a single occurrence. This rarest subset of CNVs would be expected to show enrichment under the model that genetic causes of schizophrenia are individually unique in some proportion of patients. Indeed, this set of CNVs showed a 1.45-fold increase in cases (empirical $P = 5 \times 10^{-6}$). On average, 13.1% of cases of schizophrenia possessed a deletion or duplication observed only once in the sample, in contrast to 10.4% of controls. Under a model in which very rare (occurring in under 1/1,000 individuals) inherited or recurrently de novo events increase risk, we would expect to observe a greater overall burden in schizophrenia. Although our study was statistically underpowered to identify the actual loci involved, such variants could in theory be mapped in extremely large samples. In this intermediate group, we observed 2,465 CNVs occurring between two and six times in the total sample, for which there was an increased burden, both for number of CNVs (empirical P = 0.0013) and gene count (empirical $P = 5 \times 10^{-4}$).

Because several known genomic disorders of the nervous system result from large CNVs, which are often many hundreds of kilobases long¹¹, we additionally stratified by size of event (Table 3). Of deletions, only larger (more than 500 kb) variants were enriched (empirical $P = 3 \times 10^{-4}$) despite being the least frequent set of CNVs (n = 285), displaying a 3.57-fold increase in gene count between cases and controls (empirical $P = 2 \times 10^{-5}$). In contrast, shorter duplications showed a stronger association with disease than longer duplications, albeit with a smaller fold increase than deletions (Table 3).

In general, the gene count definition of CNV burden yielded stronger results, particularly for deletions (gene count $P = 3 \times 10^{-5}$ versus number P = 0.11; Table 2). In fact, dividing all CNVs into two sets, of those that intersect at least one gene and those that do not, we saw an increased burden only in the number of 'genic' CNVs ($P = 5 \times 10^{-6}$; Supplementary Table 2) and not for non-genic CNVs (P = 0.16). There was a similar trend for CNVs seen two to six times when comparing enrichment in genic and non-genic CNVs ($P = 7 \times 10^{-4}$ and 0.19, respectively) but not single-occurrence CNVs ($P = 6 \times 10^{-4}$ and 6×10^{-4} , respectively). These results may reflect biological distinctions, although they may to some extent also reflect variable performance in CNV detection for different classes of variant.

We conducted a set of analyses to rule out several sources of bias and confounding in the primary genome-wide burden analysis (Supplementary Tables 3-6). Although, in general, low specificity and sensitivity decrease power, of concern here is potential measurement error that varied systematically between cases and controls, leading to spurious results. In this respect, an obvious concern is that both Affymetrix 5.0 and 6.0 arrays were used; as a consequence, we performed all analyses controlling for array type. As described in Supplementary Information, the primary result was also robust to the following. First, in addition to array type, we controlled for sample collection site, genotyping plate and average probe variance. Second, sensitivity analyses showed that no single sample collection site accounted for the observations. Third, we restricted analysis to the most homogeneous 90% of the sample with respect to intraindividual probe variance. Fourth, if differences in CNV burden between cases and controls were purely due to unmeasured confounders, we would not expect an enriched gene count after controlling for the overall extent and rate of CNVs. However, after controlling for overall (genic and non-genic) CNV burden there remained a significantly enriched gene-count burden in patients with schizophrenia.

Our large sample size further enabled us to search for specific CNV regions associated with schizophrenia. One locus previously reported to increase risk for schizophrenia is 22q11.2 (17–21 megabases (Mb)), at which hemizygosity occurs in 1 in every 4,000 live births¹². These deletions produce a range of clinically heterogeneous phenotypes, including velo-cardio-facial syndrome and DiGeorge syndrome, that

	Size range (kb)			CNV burden (numbe	er)	CNV burden (gene count)		
CNV type		CNV (n)	Р	Case/control ratio	Baseline rate (controls)	Р	Case/control ratio	Baseline rate (controls)
Deletions and duplications	100 - 200	3,725	0.0017	1.15	0.55	8×10^{-6}	1.35	0.73
	200 - 500	2,156	0.028	1.11	0.32	0.0088	1.25	0.66
	≥500	872	0.0013	1.32	0.12	8×10^{-4}	1.79	0.62
Deletions only	100 - 200	1,612	0.54	1.02	0.25	0.28	1.07	0.26
	200 - 500	755	0.39	1.04	0.12	0.059	1.27	0.14
	≥500	285	3×10^{-4}	1.67	0.03	2×10^{-5}	3.57	0.14
Duplications only	100 - 200	2,113	10^{-4}	1.26	0.30	2×10^{-6}	1.50	0.47
	200 - 500	1,401	0.017	1.14	0.20	0.026	1.24	0.52
	≥500	587	0.11	1.17	0.09	0.17	1.29	0.48

The table shows an analysis of global CNV burden in cases versus controls. CNVs were stratified into three size categories (100–200 kb, 200–500 kb, and 500 kb or more). See Table 2 for further details.

together are known as 22q11.2 deletion syndrome (22q11.2DS)¹²; about 30% of carriers develop psychosis¹². Previous studies estimated the frequency of 22q11.2 deletions to be 0.6-2.0% in cases of schizophrenia, although many of these studies had technically incomplete characterization of this region¹⁹. We therefore expected to find examples of 22q11.2 deletions in our sample of 6,572 individuals. The most common form of 22q11.2DS is a 3-Mb loss (about 90% frequency), although a nested 1.5-Mb deletion is also observed (about 7%) along with infrequent (about 3%) atypical deletions²⁰. We identified 13 large deletions (more than 500 kb) in cases of schizophrenia within this interval, and none in controls (Supplementary Table 7). Of these, six were consistent with the larger deletion, five were consistent with the shorter deletion, and two were atypical. The 11 samples with typical deletions defined an interval with the strongest association (empirical P = 0.0017; genome-wide corrected P = 0.0046; odds ratio = 21.6) (Fig. 1a). Controlling for sample collection site



Figure 1 Regions with excess large deletions in cases. a, The positions of CNVs of more than 500 kb across the chromosome 22q11.2 region. Red lines, case deletions; horizontal dashed sections, qPCR or visual inspection of array intensity data suggest an extended deletion; green lines, locations of low-copy repeats (LCR22-2–LCR22-4).; grey lines, recurrent 3-Mb and 1.5-Mb velo-cardio-facial syndrome (VCFS) deletions. qPCR primers are marked by vertical dashed lines. **b**, Chromosome 15q13.3 region, as above, except for the locations of breakpoint regions (BP4 and BP5; green) and the critical region defined previously¹⁷ (grey line). **c**, Chromosome 1q21.1, as above, except that a single deletion identified in a control subject is marked by blue line. Genes based on build 35 UCSC browser (Supplementary Table 8).

or genotyping plate instead of array type did not change the results (Supplementary Table 10). The two other atypical deletions in this region overlap the distal end of the 3-Mb variant. Deletion events within the region were confirmed in all 13 patients by quantitative polymerase chain reaction (qPCR) with three individual assays (Supplementary Fig. 1 and Supplementary Tables 11 and 12). Our findings provide additional evidence that hemizygosity in 22q11.2 is a rare but powerful risk factor for schizophrenia.

The larger 22q11.2 deletion harbours 43 genes (Supplementary Table 8). Despite the efforts of many groups, the psychiatric symptoms observed in 22q11.2DS have not been ascribed to a reduced copy number of any individual gene¹². Variants within *COMT*, which encodes catechol-*O*-methyltransferase, an enzyme responsible for degrading catecholamines, including dopamine, have been implicated in a wide variety of phenotypes, but with inconsistent results¹².

Removing the thirteen 22q11.2DS individuals, we observed a further 271 deletions of more than 500 kb (161 in cases and 110 in controls). Two additional regions (15q13.3 and 1q21.1) were identified that harboured a significant excess of deletions in cases of schizophrenia after correction for multiple testing (Fig. 1b, c; see Supplementary Table 7 for case descriptions). On chromosome 15 (28–31 Mb) there were deletions in nine cases and no controls (empirical P = 0.0029; genome-wide corrected P = 0.046; odds ratio 17.9). On chromosome 1 (142.5-145.5 Mb) there were ten deletions in cases and one in controls (empirical P = 0.0076; genome-wide corrected P = 0.046; odds ratio 6.6). All 20 large deletions at 15q13.3 and 1q21.1 were validated by one or more qPCR reactions (Supplementary Fig. 1 and Supplementary Tables 11 and 12). The multiple test correction factors were small as a consequence of our having restricted attention to this small class of rare variants. We did not observe any regions with a corrected P < 0.05 for either duplications or smaller (less than 500 kb) deletions. In addition, the primary CNV burden tests remained significant after removing individuals with a deletion at one of these three loci (number $P = 10^{-4}$ and gene count $P = 3 \times 10^{-5}$); for deletions of more than 500 kb specifically, the burden test remained significant for number (P = 0.02) but not for gene count (P = 0.11).

The large deletions on chromosome 15q13.3 have not previously been associated with schizophrenia. This region does not include the nearby critical region for Prader–Willi/Angelman syndrome²¹ but is consistent with the critical region defined by recurrent deletion in cases of mental retardation with seizures that have been reported recently¹⁷. Furthermore, our estimated breakpoints fall within the segmental duplications reported (BP4 and BP5). In the present study, evidence consistent with mildly impaired cognition was seen in five of the nine patients with deletions, and one individual also had a history of epilepsy (Supplementary Table 7). This broad region has been the focus of previous genetic studies in schizophrenia. The gene *CHRNA7*, encoding the α_7 subunit of the nicotinic acetylcholine receptor, is a candidate based on an initial identification from linkage analysis of auditory evoked potential deficits observed in patients with schizophrenia^{22,23}.

The deleted region on 1q21.1 is consistent with a previously reported *de novo* deletion in a patient with learning disability and seizures²⁴ and two patients with autism (one *de novo* and one inherited)¹⁰. In the present study, three cases had mild cognitive abnormalities and one had a history of epilepsy (Supplementary Table 7). The region contains 27 known genes, most of which are expressed in the brain (Supplementary Table 8), and previous reports have shown linkage^{25,26} but there have been no previous reports of CNVs associated with schizophrenia.

Regions of highly homologous segmental duplication flank the deletions we report at 22q11.2, 15q13.3 and 1q21.1. A prominent mechanism for CNV genesis is non-allelic homologous recombination mediated by segmental duplications, resulting in deletions and reciprocal duplications of the interval between segmental duplications^{16,27}. Neurodevelopmental and psychiatric syndromes have been associated with deletions and duplications flanked by segmental duplications, many of which occur *de novo*^{10,11,17}. Segmental duplications

and non-allelic homologous recombination mediate CNVs at 22q11.2 (ref. 28) and may be involved in the genesis of CNVs at 15q13.3 (ref. 17) and 1q21.1, although other mechanisms may be involved²⁹.

While this work was under review, Walsh et al.² reported a higher frequency of cases with CNVs (15%, based on 23 CNVs in 150 patients with schizophrenia) than in controls (5%). Of the 21 autosomal case CNVs identified in that report, we observed overlapping control CNVs at six loci (for example DLG2 and PTPRM; Supplementary Table 9), illustrating that large sample numbers are needed to conclude that any one particular CNV or implicated gene can cause schizophrenia. Our global burden analysis demonstrated that, on aggregate, single-occurrence and very rare (under about 1 in 1,000) CNVs have increased rates in cases of schizophrenia in comparison with controls, in line with Walsh et al.². This indicates that at least some of these rare CNVs seen in cases but not in controls are probably risk factors for schizophrenia, although like Walsh et al. we are unable to identify which. Some examples of possible risk CNVs that were observed multiple times only in cases include deletions at 12p11.23 (four cases) and 16p12.2-12.1 (four cases). These deletions were more than 500 kb, flanked by segmental duplications and spanning several brain-expressed genes. In addition, duplications in two genes relevant to neural development and growth (NOTCH1 and p21-activated kinase 7, PAK7) were found in five and six cases, respectively, and no controls. Furthermore, we identified CNVs at two recently reported loci, NRXN1 and CNTNAP2 (refs 3, 5) (Supplementary Fig. 2).

The aetiology of schizophrenia has been vigorously debated. We now have strong and replicated² evidence that individuals with schizophrenia have a greater burden of structural variation across their genomes. Our data show that CNVs in at least three loci act as strong risk factors for schizophrenia in a minority of individuals. We can therefore now posit that some cases of schizophrenia are 'genomic disorders'¹⁶ although we do not yet know whether the risk is specific for schizophrenia as opposed to a more general risk factor for neuropsychiatric or central nervous system illness.

Exactly how a subtle, 1.15-fold increase in CNV burden translates mechanistically into illness in a given patient is currently unknown. We also do not know whether common genetic variants of more subtle effect are components of the aetiology of schizophrenia, an empirical question that we and others are addressing. Similarly, we do not know how environmental risk or protective factors might act in concert with specific CNVs or with the overall burden of CNVs.

A critically important goal will be to determine the full clinical and phenotypic spectrum in carriers of these deletions. Our data provide preliminary evidence of a variable phenotype in patients with schizophrenia who would otherwise be regarded as clinically typical. Examining the role of these variants in related psychotic disorders, such as bipolar disorder, is imperative. Further work explicating the epidemiology and mechanism of these variants in schizophrenia may ultimately lead to a role for them in genetic counselling and understanding disease biology.

Note added in proof: Samples from the University of Aberdeen were genotyped independently by the SGENE Consortium³⁰.

METHODS SUMMARY

Cases satisfied DSM-IV³¹ or ICD-10 (ref. 32) criteria for schizophrenia and were broadly representative of clinical cases in contact with psychiatric services. DNA was extracted from whole blood, with approval from institutional review boards. CNVs were identified with the Birdseye package¹⁵ and analysed with PLINK v1.03 (ref. 14). See Supplementary Information for details. A list of all CNVs passing quality control is available (http://pngu.mgh.harvard.edu/isc/).

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Supplementary Information is linked to the online version of the paper at www.nature.com/nature.

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The International Schizophrenia Consortium

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